October 5, 2018

Dear Laboratory Director,

Carbapenem-resistant *Enterobacteriaceae* (CRE) and carbapenemase-producing CRE (CP-CRE) are a concerning public health threat due to multidrug-resistance, ability to cause serious invasive infection, and potential for rapid spread.

On July 30, 2018, the Georgia Department of Public Health (DPH) updated the Notifiable Disease/Condition Reporting List to require reporting of positive CRE laboratory results to DPH and submission of isolates to the Georgia Public Health Laboratory (GPHL).

CRE Case Definition:

Enterobacter species, Escherichia coli or Klebsiella species, from any clinical specimen (including screening/surveillance cultures) and resistant to at least one carbapenem (minimum inhibitory concentrations of \geq 4 mcg/ml for meropenem, imipenem, and doripenem or \geq 2 mcg/ml for ertapenem) **OR** production of a carbapenemase (e.g., KPC, NDM, VIM, IMP, OXA-48 carbapenemase) demonstrated by a recognized test (e.g., polymerase chain reaction, metallo- β -lactamase test, modified Hodge test, Carba NP).

Current CLSI (June 2010) carbapenem breakpoints must be used to ensure CRE are properly identified.

Include susceptibility results for all antibiotics tested (quantitative MIC value, and qualitative interpretation [S, I, R]), plus all results regarding carbapenemase production (positive or negative). Include submission of isolates meeting this surveillance definition to GPHL.

Three attached resources provide information needed to report and submit CRE:

- 1. CRE Reporting Guidelines (pages 2-7)
- 2. CRE Isolate Submission Guidelines (page 8)
- 3. CRE-Specific GPHL Isolate Submission Form (page 9; additional copies available here: https://dph.georgia.gov/notifiable-hai-reporting)

We thank you for your cooperation in our initiative to better understand and control CRE in Georgia.

Sincerely,

Cherie L. Drenzek, MS, DVM State Epidemiologist



Carbapenem-Resistant *Enterobacteriaceae* (CRE) Reporting Guidelines

What to Report

Organism

- 1. *Enterobacter* species, *Escherichia coli*, or *Klebsiella* species resistant to imipenem, meropenem, doripenem, or ertapenem isolated from any body site or from clinical and surveillance specimens
- 2. *Enterobacter* species, *Escherichia coli* or *Klebsiella* species that demonstrate production of a carbapenemase (e.g., KPC, NDM, VIM, IMP, OXA-48) by a recognized test (e.g., polymerase chain reaction, metallo-ß- lactamase test, modified Hodge test, Carba NP, carbapenemase inactivation method [CIM] or modified CIM)
- 3. Any diagnosis of CRE or CP-CRE infection or colonization

Carbapenem Susceptibility Results

The current CLSI MIC breakpoints for carbapenems are listed below:

Carbanenem	MIC Breakpoints (µg/mL)				
Carbapeneni	Susceptible	Intermediate	Resistant		
Ertapenem	≤0.5	1	≥2		
Doripenem	≤1	2	≥4		
Imipenem	≤1	2	≥4		
Meropenem	≤1	2	≥4		

Patient and Specimen Variables

The following information is to be reported by the laboratory:

- <u>Patient information</u>: name, medical record number, date of birth, gender, race, ethnicity, address, city, state, county, and phone number
- <u>Specimen information</u>: organism name (genus and species), specimen source, **all antimicrobial susceptibility results** (including agent tested, testing method, MIC values, and MIC interpretation as S, I, or R), and carbapenemase testing results (including test method and results)

Unlike other notifiable conditions, **antimicrobial susceptibility results are required** for a report of CRE to be considered complete.



How to Report

Results are to be reported using Electronic Laboratory Reporting (ELR). If your laboratory is currently reporting to DPH using ELR, you may require modifications to meet the required reporting for CRE. We recommend that you work with the Information Technology staff at DPH to ensure that you can report the fields listed under "What to Report" above.

To test your ability to send CRE messages, send a test message through the existing PHINMS/SFTP connection that you use, ensuring that the name of the file is different than production ELR data. For example, if the name a facility sends for production ELR is "MYHOSPITAL_ELR_"+date, then sending as 'TEST_CRE_'+date will ensure that the file will not get picked up and processed. Once the message has been sent, send an email to Patrick Pitcher (patrick.pitcher@dph.ga.gov) letting him know that a file is ready for him to test (please include the facility name for which you are testing along with the keyword "CRE" in the subject line of email).

If you are new to ELR reporting, register at this address: <u>https://dph.georgia.gov/meaningful-use</u>

LOINC and SNOMED Codes

The following list of test/result codes can be used for ELR reporting and creating result queries:

LOINC	LOINC Name	SNOMED	SNOMED Name		
11475-1		112283007	Escherichia coli		
	Microorganiem identified, Drld, DT, www. Nom.	1485002	Enterobacter cloacae		
	Culture	62592009	Enterobacter aerogenes		
	Culture	56415008	Klebsiella pneumonia		
		40886007	Klebsiella oxytoca		
75757-6		112283007	Escherichia coli		
		1485002	Enterobacter cloacae		
	Bacteria identified in Isolate by MS.MALDI-TOF	62592009	Enterobacter aerogenes		
		56415008	Klebsiella pneumonia		
		40886007	Klebsiella oxytoca		

Organism Identification

Susceptibility Information

LOINC	LOINC Name	SNOMED	SNOMED Name
56031-8	Doripenem [Susceptibility] by Minimum		
	inhibitory concentration (MIC)		
35801-0	Ertapenem [Susceptibility] by Minimum		
	inhibitory concentration (MIC)		
279-0	Imipenem [Susceptibility] by Minimum inhibitory		
	concentration (MIC)		
6652-2	Meropenem [Susceptibility] by Minimum		
	inhibitory concentration (MIC)		



Carbapenemase Detection Methods

LOINC	LOINC Name	SNOMED	SNOMED Name		
86930-5	Carbananamasa [Procense] in Isalata	10828004	Positive		
	Carbapenemase [Presence] in isolate	260385009	Negative		
		10828004	Positive		
74676-8	Carbananamasa [Tuna] in Isalata hu Carba ND	260385009	Negative		
	Carbapenemase [Type] In Isolate by Carba NP	42425007	Equivocal		
		11896004	Intermediate		
	Carbananamasa Datastad via Carbananam	10828004	Positive		
	Lipactivation Method (CIM)	260385009	Negative		
		42425007	Equivocal		
		10828004	Positive		
	Carbapenemase Detected via Modified	260385009	Negative		
	Carbapenem Inactivation Method (mCIM)	42425007	Equivocal		
		11896004	Intermediate		
85502-3	Carbapenemase resistance genes panel -	10828004	Positive		
	Anorectal or isolate by NAA with probe detection	260385009	Negative		

Carbapenemase Genes

LOINC	LOINC Name	SNOMED	SNOMED Name
75683-3	Pla(KDC) OL Prb Mag	10828004	Positive
	BIA(RPC) QI PID WAg	260385009	Negative
75696 6	Pla(IMP) OI Drb Mag	10828004	Positive
73080-0		260385009	Negative
756841-1	Pla(NDM) OLDrb Mag	10828004	Positive
		260385009	Negative
75685 8	RIa(V/INA) OI Prb Mag	10828004	Positive
73083-8		260385009	Negative
75685 5	$R_{12}(OYA) \cap I$ Brb Mag	10828004	Positive
73083-3		260385009	Negative
	Carbapenem resistance blaIMP gene [Presence]	10828004	Positive
85498-4	in Anorectal or isolate by NAA with probe	260385009	Negative
	detection		
	Carbapenem resistance blaKPC gene [Presence]	10828004	Positive
85499-2	in Anorectal or isolate by NAA with probe detection	260385009	Negative
	Carbapenem resistance blaNDM gene [Presence]	10828004	Positive
85500-7	in Anorectal or isolate by NAA with probe detection	260385009	Negative
	Carbapenem resistance blaOXA-48 gene	10828004	Positive
85503-1	[Presence] in Anorectal or isolate by NAA with probe detection	260385009	Negative
85501-5	Carbapenem resistance blaVIM gene [Presence]	10828004	Positive
	in Anorectal or isolate by NAA with probe detection	260385009	Negative



Parent/Child ELR Relationship for Culture and Susceptibility Testing

Modified from the Tennessee Department of Health ELR On-Boarding Handbook

To fully assess antimicrobial resistance, DPH needs to receive enough information about resistance testing for specific organisms. This includes:

- 1. The antimicrobial/bactericidal agent being tested
- 2. The method of testing (Kirby-Bauer, MIC, etc.)
- 3. The actual quantitative and qualitative results and interpretations

Specific fields in the HL7 message allow for the antimicrobial susceptibilities to be reported to public health. The messages should contain the organism, antibiotic susceptibilities, and the specimen source. The parent observation is the identified observation and the child observation is the antibiotic susceptibility result. The child observations should list all antibiotics tested against the organism, the measured MIC values, and the phenotypic interpretation.

How to link parent-child observations using HL7 2.5.1:

MSH PID ORC OBR|1|Placer|Filler OBX|1|Observation^Identifier|ObservationSubID|ObservationValue SPM OBR|2||||||||||||||||Observation&Identifier^ObservationSubID^ObservationValue|||Placer^Filler OBX|1 OBX|2 OBX|3 SPM

Example [OBR-26] Parent Result:

[600-7&Microorganism identified&LN&CULT&Culture&L^1^Streptococcus pneumoniae]

- The first component <600-7&Microorganism identified&LN&CULT&Culture&L> consists of the test codes and descriptions for a microbial culture that appeared in the parent observation [OBX-3].
- The second component <1> is the sub-ID in the parent organism [OBX-4].
- The third component *<Streptococcus pneumoniae>* is the result description of the parent observation. The result description should come from [OBX-5.2] of the parent observation, but may come from [OBX-5.5] if [OBX-5.2] is empty.

Additional information and message examples can be found in Appendix A of the HL7 Version 2.5.1 Implementation Guide: Electronic Laboratory Reporting to Public Health Release 1 (US Realm), with errata.



Georgia Department of Public Health | October 2018 | Page 5 of 9

Sample Message for Parent/Child ELR Relationship for Culture and Susceptibility Testing

Modified from the CSTE AR/ELR Working Group Recommended Best Practices for Surveillance of Antimicrobial Resistance via Electronic Laboratory Reporting

In the observation request (OBR) segment, the parent filler order number (highlighted in green) located in OBR 3 is linked to the child parent sequence located in OBR 29.2 to link parent culture results to child susceptibility results.

MSH|^~\&|NIST^2.16.840.1.113883.3.72.5.20^ISO|NIST^2.16.840.1.113883.3.72.5.21^ISO|NIST^2. 16.840.1.113883.3.72.5.22^ISO|NIST^2.16.840.1.113883.3.72.5.23^ISO|20120821140551-0500||ORU^R01^ORU_R01|NIST-ELR-004.01|T|2.5.1|||NE|NE|||||PHLabReport-NoAck^HL7^2.16.840.1.113883.9.11^ISO SFT|NIST Lab, Inc.^L^^^NIST&2.16.840.1.113883.3.987.1&ISO^XX^^123544|3.6.23|A-1 Lab System|6742873-12||20100617

PID|1||PATID1234^^&2.16.840.1.113883.3.72.5.24&ISO^MR^Seminole Cnty Hlth C&2.16.840.1.113883.3.0&ISO||Jones^William^A^^^L||19610615|M||2106-3^White^CDCREC|1955 Seminole Lane^^Oveido^FL^32765^USA^H^12059||^PRN^PH^1^407^2351234||||||||N^Not Hispanic or Latino^HL70189^NL^not latino^L^2.5.1

ORC|RE|ORD723222-4^2.16.840.1.113883.3.72.5.24^ISO|R-783274-4^LIS^2.16.840.1.113883.3.72.5.25^ISO||||||||57422^RADON^NICHOLAS^^Dr.^^NPI&2.16.840 .1.113883.4.6&ISO^L^^NPI||^PRN^PH^^407^2341212||||||Seminole County Health Clinic|555 Orange Ave^^Oviedo^FL^32765^B|^WPN^PH^^813^8847284|555 Orange Ave^^Oviedo^FL^32765^B

OBR|1|ORD723222-4^2.16.840.1.113883.3.72.5.24^ISO|R-783274-4 ^LIS^2.16.840.1.113883.3.72.5.25^ISO|625-4^Bacteria identified in Stool by Culture^LN^3456543^CULTURE STOOL^99USI^2.40|||20110528||||||||57422^RADON^NICHOLAS^^Dr.^^NPI&2.16.840.1.113 883.4.6&ISO^L^^^NPI|^PRN^PH^^407^2341212|||||201106010900-0500|||F

OBX|1|CWE|625-4^Bacteria identified in Stool by Culture^LN^Bacteria identified^Bacteria identified^99USI^2.40|1|85729005^Shigella flexneri^SCT^^^^Shigella flexneri|||||F||20110528||||20110531130655-0500|||Seminole County Health Department Laboratory^^^&2.16.840.1.113883.3.72.5.30.1&ISO^XX^^987|6756 Florida Avenue^Oveido^FL^32765^B|10092^Pafford^Hamlin^^^&2.16.840.1.113883.3.72.5.30.1&ISO ^L^^NPI

SPM|1|^ORD723222-4&&2.16.840.1.113883.3.72.5.24&ISO||119339001^Stool specimen^SCT^^^07/31/2012|||||||||||20110528|20110529



OBR|2||R-783274-5^LIS^2.16.840.1.113883.3.72.5.25^ISO|50545-3^Bacterial susceptibility panel in Isolate by Minimum inhibitory concentration (MIC)^LN^Bact suscept^Bacteria susceptibility^99USI^2.40||20110528||||||57422^RADON^NICHOLAS^^Dr.^^NPI&2.16.840. 1.113883.4.6&ISO^L^^NPI|^PRN^PH^^407^2341212|||201106010900-0500||F|625-4&Bacteria identified in Stool by Culture&LN&Bacteria identified&Bacteria identified&99USI^Shigella flexneri|||^R-783274-4&LIS&2.16.840.1.113883.3.72.5.25&ISO

OBX|1|SN|20-8^Amoxicillin+Clavulanate [Susceptibility] by Minimum inhibitory concentration (MIC)^LN^AmoxClav^Amoxicillin-clavulanic acid^99USI^2.40||=^16|ug/mL^microgram per milliliter^UCUM^^^1.8.2||I^Intermediate^HL70078^^^2.5.1|||F|||20110528|||||20110601090 0-0500||||Seminole County Health Department Laboratory^^^^&2.16.840.1.113883.3.72.5.30.1&ISO^XX^^987|6756 Florida

Avenue^^Oveido^FL^32765^^B|10092^Pafford^Hamlin^^^^&2.16.840.1.113883.3.72.5.30.1&ISO ^L^^^NPI

OBX|2|SN|516-5^Trimethoprim+Sulfamethoxazole [Susceptibility] by Minimum inhibitory concentration (MIC)^LN^TMP-SMX^Trimethoprim-

sulfamethoxazole^99USI^2.40||=^8^/^152|ug/mL^microgram per

milliliter^UCUM^^^1.8.2||R^Resistant^HL70078^^^2.5.1|||F|||20110528|||||201106010900-0500||||Seminole County Health Department

Laboratory^^^^&2.16.840.1.113883.3.72.5.30.1&ISO^XX^^987|6756 Florida

Avenue^^Oveido^FL^32765^^B|10092^Pafford^Hamlin^^^^&2.16.840.1.113883.3.72.5.30.1&ISO ^L^^^NPI

OBX|3|SN|185-9^Ciprofloxacin [Susceptibility] by Minimum inhibitory concentration (MIC)^LN^CIPROFLOXACIN^CIPROFLOXACIN^99USI^2.40||<=^0.06|ug/mL^microgram per milliliter^UCUM^^^1.8.2||S^Susceptible^HL70078^^^2.5.1|||F|||20110528|||||201106010900 -0500||||Seminole County Health Department Laboratory^^^^&2.16.840.1.113883.3.72.5.30.1&ISO^XX^^987|6756 Florida Avenue^^Oveido^FL^32765^B|10092^Pafford^Hamlin^^^&2.16.840.1.113883.3.72.5.30.1&ISO ^L^^^NPI

Additional Guidance

The HL7 Version 2.5.1 Implementation Guide can be found at <u>http://www.hl7.org/implement/standards/product_brief.cfm?product_id=98</u>

THE CSTE ELR Best Practices for reporting CRE can be found at https://www.cste2.org/Publications/CRE_ELR_Best_Practices_FINALv1.0_20170515.pdf

For laboratories that are currently unable to report using ELR, please contact Liz Smith, Acute Disease Epidemiology Section, for interim guidance on reporting (<u>Elizabeth.Smith@dph.ga.gov</u>).



Carbapenem-Resistant *Enterobacteriaceae* (CRE) Isolate Submission Guidelines

What are the specimen requirements?

Submit all isolates of carbapenem-resistant *Enterobacter spp*, *Klebsiella spp*, and *Escherichia coli* from any body site as pure, low passage culture on a non-inhibitory, non-selective agar slant within seven days of identification to the Georgia Public Health Laboratory. Each tube must be labeled with the patient's name, date of birth, and date of isolation.

What forms are needed?

Complete a CRE-specific GPHL form for each isolate (see next page). The form can also be found on the Georgia DPH Healthcare-Associated Infections website here: <u>https://dph.georgia.gov/notifiable-hai-reporting</u>.

How do I ship specimens to GPHL?

Triple contain isolates according to Category B UN3373 shipping regulations for infectious substances/diagnostic specimens by (1) wrapping each specimen tube in absorbent material, then (2) placing into the inner mailing container with cap securely tightened, and (3) placing into outer mailing container with completed forms. Clearly label the outer mailing container with your name/address and GPHL's address:

Georgia Public Health Laboratory 1749 Clairmont Road Decatur, Georgia, 30033

For more information see:

https://dph.georgia.gov/sites/dph.georgia.gov/files/related_files/site_page/GPHL%20PS%20Plan-8%2010%202018-Updated.pdf

What will GPHL do with the specimens I submit?

GPHL will perform additional testing to identify carbapenemase-producing CRE, to understand the epidemiology of CRE across Georgia, to aid in outbreak investigation, and to support CRE prevention initiatives.

The results of additional testing will be sent to you via mail or will be available electronically via the GPHL Web Portal. If you are not signed up for the GPHL Web Portal and would like to be, contact the DPH IT Help Desk at <u>dphall.it@dph.ga.gov.</u>

What if I participate in the Emerging Infections Program (EIP)?

If you suspect you have a cluster or outbreak of CRE or other multi-drug resistant organism, contact the DPH HAI program via email (dph.hai@dph.ga.gov) or ask for an HAI team member at 404-657-2588. Otherwise, continue your regular submission to EIP, which will forward isolates to GPHL.





GEORGIA PUBLIC HEALTH CRE/ARLN SENTINEL LABORATORY SUBMISSION FORM

Laboratory use only

HEALTH CARE PROVIDER INFORMATION PATIENT INFORMATION											
Submitter Code			Patient ID	PATIE							
Submitter Code		Number*	Last*		First*	First*		Suffix			
Submitter Name*			County of Res	idence*			DOB*				
							//				
Street Address			Home Phone:	Home Phone: Cell Phone:			Work Ph	one:			
			Address			City,	Sta	te	Zip		
City	State	Zip									
		•									
Responsible Official			EIP Number/S	pecimen	ID						
Responsible Official (Copy	To)		Race			Ethnicity		Gend	der*		
			American Ind	ian/Alaska	a Native	Hispanic	or	□ M	ale		
			Asian			Latino	Female				
				1-Americar iian/Pacif	n ic Islander	Latino					
Contact Information (Alert)	(alues)		White/ Cauc	asian		Pregnant	Pregnant?				
	ulues)		🔲 Multi-Racial			Yes	ΠNο	ΠN	/A		
			Travel in the past month? 🗌 Yes 🗌 No Travel Dates?								
EPI-14-14-4242730		Where?									
Other lab tests performed and res	ults:										
Suspected organism:											
SPECIMEN INFORMATION					•	TEST	REQUEST	ED			
Specimen Type:		0	Date of Collection*		CRE Surveillan	ce and Respo	onse				
□ Isolate (Microbial)			// X12100 M			crobial Identification for CRE CRPA					
Source*					ward to TN ARLN						
P/			Cand			ndida auris r	ida auris rule out nol Poporting (ESBL Candida				
SI			hipped:	hipped:			Canald	canalaa			
		Frozen									
		」Refrigerated ☐ Room Temperature									
S		Koom remperature									
_											
			Date of onset								
	//										
CRE submission form for Sentinel	Providers										

Last edited: April 2018